

## Nanotrap® Microbiome A; 35 mL Manual Protocol with Promega Maxwell® HT Environmental TNA Kit

**Objective:** This protocol uses Nanotrap Microbiome A Particles and Nanotrap Enhancement Reagent 2 to capture and concentrate microbes in environmental water samples. It is optimized for microbe capture from 35 mL samples and is compatible with Promega Maxwell® HT Environmental TNA Kit

### Materials and equipment:

| Sample Type  |  |
|--|--|
| Environmental Water Samples                        |  |
| Concentration Reagent                              | Vendor                                   |
| Nanotrap® Microbiome A Particles                   | Ceres Nanosciences; SKU# 44202           |
| Nanotrap® Enhancement Reagent 2 (ER2) <sup>1</sup> | Ceres Nanosciences; SKU# 10112           |
| Extraction Kit                                     | Vendor                                   |
| Promega Maxwell® HT Environmental TNA Kit          | Promega; Cat# AX9190                     |
| Materials/Equipment                                | Vendor                                   |
| Heat Block   | Southern Labware; SKUBSH200              |
| Mini Centrifuge                                    | Scientific Industries; SKU WZ-MF6000     |
| DynaMag™-50 Magnet                                 | Thermo Fisher Scientific; Cat# 12302D    |
| DynaMag™-2 Magnet                                  | Thermo Fisher Scientific; Cat# 12321D    |
| 50 mL Conical Centrifuge Tubes                     | Stellar Scientific; SKU T15-100          |
| Tube Rotator                                       | Stellar Scientific; SKU BS-RTMNI-2       |
| Serological Pipettes and Controller                | Fisherbrand; Cat# 13-678-11E             |
| 2mL Micro Centrifuge tubes                         | Stellar Scientific; SKU T20-100          |
| Mini Vortex Mixer                                  | Scientific Industries; SKU SI-236        |
| General Reagents                                   | Vendor                                   |
| Ethanol  | Decon™ Laboratories Decon Labs; # 3916EA |
| Isopropanol  | Fisher Scientific; BP26184               |
| Molecular Biological Grade Water                   | Corning; Cat# 46-000-CM                  |

<sup>1</sup> Precipitate can form in ER2 if stored below room temperature. Allow ER2 to return to room temperature to dissolve the precipitate (can invert 2-3 times to help resuspend, do not heat).

## Capture and Extract Microbes using Nanotrap Microbiome Particles

### Procedure:

#### 1. Nanotrap Microbiome A Promega Manual Procedure-Part 1

1. Invert the environmental water sample 5 times to mix. Then, let it sit for 45 seconds at room temperature. (No need to wait for sample to reach room temperature before processing)
2. Add 35 mL of environmental water sample into a clean 50 mL conical tube.
3. Add 100  $\mu$ L of Nanotrap Enhancement Reagent 2 (ER2) to the sample and then invert 2 times to mix it.
4. Add 525  $\mu$ L of Nanotrap Microbiome A Particles to the sample and then invert 2 times to mix the particles.
5. Incubate samples with Nanotrap particles at room temperature for 30 minutes.

*Note: Invert every 5 minutes or use a rotator.*

6. Place the tube on a DynaMag-50 magnetic rack to separate the Nanotrap particles from the sample for 10 minutes.
7. Using a serological pipette, discard the supernatant carefully without disturbing the Nanotrap particle pellet.

*Note: Can use a P-1000 or P-200 pipette to remove any remaining supernatant from the sample (be careful to not lose any Nanotrap particles when removing supernatant).*

8. Add 1 mL of molecular grade water to the tube and re-suspend the Nanotrap particle pellet by pipetting on the walls of the conical tube, gently re-suspend until all Nanotrap particles have been completely collected.
9. Transfer the Nanotrap particle suspension to a new 2 mL microcentrifuge tube.
10. Place the 2 mL microcentrifuge tube on a DynaMag-2 magnetic rack to separate the Nanotrap particles from the sample for 2 minutes.
11. Using a P-1000 pipette, discard the supernatant carefully without disturbing the Nanotrap particle pellet.

*Note: If any small amount of liquid is still present, use a smaller pipette to remove all the supernatant from the bottom of the tube.*

12. Add 300  $\mu$ L of Cell Lysis Solution to Nanotrap particle pellet, pipette up and down until Nanotrap particles are resuspended completely.
13. Close the tube lid, incubate the samples at RT for 10 minutes.
14. Place the 2 mL microcentrifuge tube on a DynaMag-2 magnetic rack to separate the Nanotrap particles from the sample for 2 minutes.

*Note: May need to briefly centrifuge the tube (Mini Centrifuge at 2000 g for 2-5 seconds) to remove drops from inside the lid before magnetic separation.*

15. Transfer supernatant/lysate to a new 2 mL collection tube and discard the Nanotrap particles pellet.
16. Sample is now ready for Part 2.

## **2. Nanotrap Microbiome A Promega Manual Procedure-Part 2.**

1. Add 50  $\mu$ L of Alkaline Proteinase to the sample/lysate.
2. Add 400  $\mu$ L of 100% isopropanol to the sample/lysate.
3. Add 35  $\mu$ L of Magnetic Beads Resin to the sample/lysate.
4. Vortex to mix, then incubate at RT for 20 minutes.
  1. Place on shaker/rotator.

5. Place the tube on a DynaMag-2 magnetic rack to separate the magnetic beads from the sample for 2 minutes, then discard the supernatant using a pipette.

*Note: May need to briefly centrifuge the tube (Mini Centrifuge at 2000 g for 2-5 seconds) to remove drops from inside the lid before magnetic separation.*

6. Add 1000  $\mu$ L of Dilute Wash Solution to sample and re-suspend the magnetic beads using a pipette.
7. Place the tube on a DynaMag-2 magnetic rack to separate the magnetic beads from the sample for 2 minutes, then discard the supernatant.
8. Add 1000  $\mu$ L of Dilute Wash Solution to sample and re-suspend the magnetic beads using a pipette.
9. Place the tube on a DynaMag-2 magnetic rack to separate the magnetic beads from the sample for 2 minutes, then discard the supernatant.
10. Add 450  $\mu$ L of 50% EtOH to sample and re-suspend the magnetic beads using a pipette.
11. Place the tube on a DynaMag-2 magnetic rack to separate the magnetic beads from the sample for 2 minutes, then discard the supernatant by using a pipette.
12. Add 450  $\mu$ L of 50% EtOH to sample and re-suspend the magnetic beads using a pipette.
13. Place the tube on a DynaMag-2 magnetic rack to separate the magnetic beads from the sample for 2 minutes, then discard the supernatant by using a pipette.
14. Centrifuge the tube (Mini Centrifuge at 2000 g for 30 seconds).
15. Place the tube on a DynaMag-2 magnetic rack, then remove excess 50% EtOH using a smaller pipette.

16. Open caps, allow samples to air dry at room temperature for 10 minutes.
17. Add 50  $\mu$ L of Tris-HCL (pH 8.0) to re-suspend the magnetic beads and then incubate at RT for 7 minutes (close caps).

1. Place on shaker

18. Place the tube in the DynaMag-2 magnetic rack to separate the magnetic beads from the sample for 2 minutes.

*Note: May need to briefly centrifuge the tube (Mini Centrifuge at 2000 g for 2-5 seconds) to remove drops/condensation from inside the lid before magnetic separation.*

19. Transfer the supernatant to a new tube, the sample is ready for downstream analysis or can be stored at  $-80^{\circ}\text{C}$ .

*Note: Multiple freeze-thaw cycles may cause degradation.*

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